

J. Empir. Soc. Sci. Stud. 7(4)

Advancements in Sensor Technologies for Microfluidic Systems: Implications for Drug Discovery and Plant Pathogen Detection

Mei-Lin Chen

Department of Biomedical Engineering, National Chiayi University

Jian-Hua Wang

National Pingtung University

Abstract

Microfluidic systems integrated with sensors have shown great promise for applications in drug discovery and plant pathogen detection. Recent advancements in sensor technologies including electrochemical, optical, mechanical, and mass sensitivity detectors have enabled accurate, rapid, and cost-effective analysis of biochemical samples on microfluidic chips. The integration of microfluidics with sensors provides capability for high-throughput analysis, portability, reduced sample requirements, and process automation. The small dimensions of microfluidic channels allow precise manipulation of very small volumes of fluids, facilitating automation. This article reviews recent research on novel microfluidics-based sensors and their emerging implications on pharmaceutical research and agriculture. For drug discovery, microfluidic platforms with embedded sensors enable refined in vitro models that better emulate human physiology for reliable toxicity assessment and drug screening. Lab-on-chip microfluidic sensors with capabilities in nanoliter sample volumes are also cutting analysis times and costs in the discovery pipeline. High-throughput screening against large compound libraries and optimizing synergistic combinations of drug candidates is now possible. For plant pathogen screening, microfluidic immunoassays and molecular diagnostic devices facilitate rapid, sensitive and portable detection alternatives to conventional laboratory-based screening tools. Smartphone-readable microfluidic sensor strips are creating opportunities for instant field tests without requiring extensive supporting infrastructure. Future priorities focus on expanding organ and disease models, mobile diagnostics for on-site analysis, smartphone integration, process automation, training provisions, and cross-disciplinary collaborations to translate more microfluidic sensors from lab to application. Overall, microfluidics-integrated sensor technologies promise to transform drug development and agriculture landscape through early disease diagnosis, toxicology prediction, and precise interventions during disease/treatment stages.

Keywords: High-throughput screening, Microfluidics, Sensor technologies, Electrochemical sensors, Optical sensors.

Introduction

Microfluidics refers to the science of manipulating and controlling fluids at submillimeter scale utilizing channels with dimensions typically ranging from 5-500 micrometers. Operating with such minuscule volumes leads to advantages of portability, reduced reagent consumption, faster analysis, improved process control and automation [1]. The behavior of fluids at the microscale level differs significantly from that in macroscale systems due to factors like increased surface area to volume ratio, laminar flow, and dominance of surface tension over gravity. As a result, scaling down conventional lab processes to microfluidic format entails precise engineering control over device material, surface properties, geometries, energy input and fluid movement. Sophisticated and affordable microfabrication techniques originally developed for microelectronics industry enabled the emergence of microfluidics technology over the past three decades. Especially, rapid prototyping based on soft lithography to pattern microchannels in polymers provided flexibility and accessibility for diverse applications [2].

Various mechanisms have been utilized to manipulate fluids through microfluidic devices including pressure-driven flow, capillary action, centrifugation, and use of pumps or valves. Integration of functional elements like mixers, separators, concentrators and extractors within self-contained microfluidic chips facilitated automation of complex biochemical processes [3]. Furthermore, capability to shrink and connect laboratory analysis protocols composed of multiple sample handling and processing steps into an integrated "lab-on-a-chip" system paved way for micrototal-analysis systems. Such microfluidic platforms minimize cumbersome manual steps between different tasks thus reducing contamination risks, human errors and process variability yielding improved accuracy. Additionally, compartmentalization of various reagents on a single chip enable manipulating precise ratios. Various

microfluidic components for fluidic handling at nanoliter or even picoliter volumes are available [4]. For instance, microdroplet systems involving water-in-oil emulsions allow high-throughput manipulation of discrete droplets in microchannels as minute reactors. Microfluidic large scale integration borrowing manufacturing methods from electronics industry also enabled mass production of complex microfluidic circuits. Further innovations in device architecture, functional design and reproducible fabrication techniques promise continued progress in achieving programmable "flow computers" [5].

To leverage these capabilities for biochemical analysis and medical diagnostics, microfluidic devices must seamlessly combine sample handling, reactions, separations and detection modules. Effective integration relies on implementing sensors that can provide rapid, reliable and sensitive analysis of miniscule sample volumes. Recent years have witnessed remarkable growth in detection methods tailored for microfluidic integration such as optical/electrochemical transduction, mechanical microresonators, microelectronics, and recognition elements like biomolecular probes and nanoparticles [6]. For example, micropatterned electrodes yield higher signal-to-noise ratios enhancing limits of detection. Microfabrication methods allow positioning detector surfaces closer to samples further augmenting sensitivity to trace analytes. Microfluidic environments enable accelerated binding kinetics leading to rapid analysis. Additional modules for sample clean-up and target preconcentration directly on chip minimize external sample preparation improving quality. Miniaturized microfluidic sensing systems have surpassed performance of established analytical techniques in terms of reduced response time, sample volume, power consumption and increased portability enabling point-of-need analysis. Such compelling advantages enabled transition of microfluidics from academic curiosity to commercial reality with real-life applications. Major application areas which have benefited significantly from adopting microfluidic technology include medical diagnostics, pharmaceutical research, molecular biology and agriculture [7]. Specifically, microfluidics-based platforms integrated with various detection modalities have shown tremendous promise for bioanalysis applications by providing quantitative and definitive information on diverse analytes ranging from biomarkers to toxins and pathogens [8]. The following sections highlight two key emerging areas with significant recent progress in developing integrated microfluidic sensors – pharmaceutical research focusing on drug discovery/testing and agricultural applications targeting food safety/plant pathogen detection [9]. The extended introduction provides more details on fundamental microfluidics principles, evolution of technology over the years, key component integrations, advantages compared to conventional techniques and an overview of major application areas. Let me know if you need any other changes to this section!

Sensor Technologies for Microfluidic Systems

Various sensors leveraging electrical, optical, mechanical, or mass detection techniques have been integrated with microfluidic chips to meet the detection needs of different applications (Table 1).

Table 1. Sensor techniques integrated with microfluidic systems and their detection methods.

Electrochemical Sensors: Electrochemical sensors offer the advantages of rapid response, high sensitivity, and low cost which make them suitable for integration with microfluidics. Common electrochemical detection methods include potentiometry, amperometry, conductometry and capacitance measurements based on interactions between target analytes and electrodes / probes generating a detectable electrical signal proportional to analyte concentration [10]. Recent advancements in microfabrication have enabled fabrication of microelectrodes and integration with microfluidics leading to improved performance and automated high-throughput analysis.

Optical Sensors: Optical detection methods are the most prevalent techniques used in microfluidic devices as they provide high sensitivity and avoid problems related to electrode fouling in electrochemical detection [11]. Common optical detection modes include absorbance, fluorescence, chemiluminescence, surface plasma resonance (SPR) and Raman spectroscopy. Optical fibers integrated with microfluidic channels have enabled improved excitation and emission collection leading to enhanced detection limits. Moreover, advances in electromagnetic simulations, waveguide materials, surface functional chemistries, and microfabrication processes have paved the way for developing integrated photonic lab-on-chip sensors combining microfluidics with nanophotonic [12].

Mechanical Sensors: Micromechanical sensors capable of detecting ultra-small mass changes have also been combined with microfluidics for rapid biomarker analysis. These sensors consist of micron-sized cantilevers that resonate at specific frequencies and have the ability to accurately detect slight mass changes with high precision when target molecules bind on functionalized surfaces [13]. The resonance frequency shifts caused by the bound target mass provide quantitative information proportional to the concentration of biomarkers. Mechanical sensing offers the benefits of fast response time and label-free detection that are extremely useful for applications requiring rapid analysis.

Mass Sensitivity Sensors: Mass sensitive detection methods relying on electrochemical quartz crystal microbalance (EQCM), microcantilevers, magnetic nanoparticles, and carbon nanotubes have been implemented with microfluidic systems for bioanalysis applications. Compared to traditional analytical techniques, these sensing platforms integrated with microfluidics provide faster response, higher sensitivity, lower sample volume requirement, and ease of operation for rapid biomarker screening [14].

Technological advancements have enabled the development of highly sensitive sensors based on electrochemical, optical, mechanical and mass detection methods with greatly improved performance characteristics. Seamless integration of these sensors with microfluidic channels has paved the path for automated lab-on-a-chip devices with powerful capabilities for drug discovery and agricultural applications.

Emerging Trends in Sensor Technologies for Microfluidics

Current research is focused on developing sensors tailored to meet applicationspecific needs along with solutions for integrating multi-mode detection techniques to enable multiplex analysis on microfluidic chips. For instance, electrochemical sensors are being improved to allow detection of redox events from small sample volumes with enhanced sensitivity and reproducibility. Novel optical sensors are also being developed using optimized materials and excitation sources to achieve the lowest limits of detection for target analytes [15]. Additionally, considerable research efforts emphasize shrinking the sensor footprint and developing low-power detection solutions for implementing mobile, field-deployable microfluidic devices. Furthermore, machine learning approaches are being applied to improve sensor performance, optimize device fabrication protocols, and accurately analyze data from microfluidics-integrated sensors [16]. These advancements promise continued innovation in microfluidics-based sensors with significant implications for pharmaceutical research and agricultural applications [17].

Applications in Drug Discovery: The use of microfluidic devices integrated with various sensors shows significant advantages over conventional screening platforms by providing improved automation, reduced timescale and costs for drug discovery (Figure 1). This section highlights key applications of microfluidics-based sensors for steps involved in drug development including target identification, lead compound screening, optimization and toxicity testing.

Identification of Drug Targets: A crucial early step in drug discovery involves identifying and validating disease-relevant biomolecular targets that can be modulated by potential therapeutic compounds. Microfluidic devices integrated with high-throughput sensors have become invaluable for target identification by enabling rapid analysis of target-ligand interactions. For instance, SPR microfluidic sensors allow real-time, label-free detection of binding affinities and kinetics between immobilized target proteins and various ligands, facilitating highthroughput screening. Similarly, living cell-based microfluidic platforms with integrated sensors provide means for efficiently detecting functional effects of drugs on relevant molecular targets and signaling pathways inside human cells. Such cellbased assays better recapitulate in vivo microenvironments compared to simpler biochemical assays [18]. Additionally, organs-on-chips consisting of microfluidic devices lined with human organ-specific cells integrated with sensors have shown promise for understanding complex target biology and assessing efficacy of drug candidates. Overall, these sensor-integrated microfluidic devices have the potential to transform early stages of drug discovery by enabling rapid validation of targets [19].

High-Throughput Screening of Lead Compounds

Once promising targets are identified, large chemical libraries need to be screened to determine lead compounds that can effectively bind and modulate the target. Conventional screening approaches use multi-well plates which require large sample volumes and are time consuming and costly. Microfluidic platforms integrated with electrochemical, optical and mass sensitive detection methods have been used to

develop label-free, high-throughput screening systems capable of rapid analysis using extremely small sample volumes. For example, an electrochemical microfluidic device was used to screen a library of over 7000 compounds to discover inhibitors for the enzyme phosphodiesterase-5 with higher sensitivity compared to standard assays. Such microfluidics-enabled high-throughput screening systems provide significant advantages over traditional approaches. Furthermore, microfluidic microarrays patterned with different testing conditions have been utilized for rapidly optimizing parameters for lead compounds. Overall, sensorintegrated microfluidic platforms have demonstrated potential for accelerating lead discovery [20].

Lead Optimization: The hits identified from primary compound screening require further optimization to select lead compounds with maximum potency and druglikeness before clinical testing. Microfluidic devices integrated with cell-based assays and sensors facilitate lead optimization by enabling rapid analysis of absorption, distribution, metabolism and elimination (ADME) characteristics. For instance, a lung-on-a-chip model lined with human cells was used with integrated sensors to determine toxicity profiles and optimal doses for lead compounds without relying on animal models [21]. Additionally, liver-on-chip platforms have been extensively utilized for evaluating liver toxicity of drugs at early stages thereby improving success in clinical trials. Microfluidic organs-on-chips thus provide means for developing physiologically relevant models of human pharmacokinetics and toxicity. Furthermore, combination drug therapy using multiple lead compounds is emerging as an effective treatment strategy for complex diseases like cancer. However, determining optimal drug ratios for synergistic effects involves extensive experimentation. Microfluidic sensors integrated with cell culture models provide assay flexibility and sensitivity for efficiently optimizing combinatorial doses to identify rational drug cocktails. Overall, adopting microfluidics-based platforms early in the pipeline has potential to nominate better lead candidates, thereby accelerating pharmaceutical research [22]**,** [23].

Toxicity Testing: The high costs associated with late-stage drug failures due to adverse effects demands reliable toxicity testing methods early in development prior to human trials. Microfluidic platforms integrated with sensors facilitate evaluation of drug safety by providing human cell-based models that replicate key organ physiology and enable detection of various toxicity endpoints. Sensors integrated with perfusable vascular channels lined with organ-specific cells in organs-on-chips have enabled analysis of multiple toxicity parameters over weeks to reliably determine safe drug doses. Such capability for long-term, repeated-dose toxicity testing fills a technology gap that animal models could not address reliably. Additionally, exposure of human liver cells to drugs in a microfluidic device led to

improved prediction of liver injury in clinical trials compared to standard assays. Another study detected cardiotoxic effects of cancer drugs using heart-on-a-chip models that conventional cytotoxicity assays failed to capture. Furthermore, linking multiple organs on microfluidic devices enabled evaluating systemic effects of drugs on interconnected organ systems through integrated electrochemical sensors, better mimicking clinical outcomes. By providing miniaturized models of human physiology with functional readouts, organs-on-chips integrated with sensors are transforming toxicity testing in pharmaceutical research [24].

Table 2 highlights key examples demonstrating applications of microfluidicsintegrated sensors for overcoming limitations in various drug discovery stages from identifying molecular targets to clinical translational research, quintessentially transforming the development pipeline.

Table 2. Examples of microfluidics-sensor integration advancing different drug discovery phases

Applications in Plant Pathogen Detection

Rapid, robust and automated pathogen detection tools are vital for agriculture to facilitate early intervention, inform breeding programs for enhancing host resistance and guide application of chemicals [25]. Conventional techniques for plant pathogen detection rely on immunoassays, polymerase chain reaction (PCR) or culturing, which can be time-consuming, labor-intensive and require centralized laboratory infrastructure. Recent research has focused on developing portable, rapid detection methods by integrating microfluidic devices with various detection modalities to meet the needs for agricultural applications. This section describes applications of microfluidics-based sensors for plant pathogen screening and opportunities for mobile, on-site testing to enhance food safety and security [26].

Microfluidic Immunoassays: Immunoassay methods leveraging the specific binding between antigens and antibodies have been extensively used for detecting plant pathogens. Conventional immunoassays rely on enzyme-linked immunosorbent assay (ELISA) performed in 96 well plates. Microfluidic immunosensors dramatically lower analysis time and enhance sensitivity compared to traditional ELISA. For example, an electrochemical immunosensor integrated with a microfluidic chip detected Phytophthora infesting causing the late blight disease in potatoes with 4-fold higher sensitivity and 8-fold faster analysis compared to standard ELISA. Similarly, a microfluidic magnetoelastic immuno-device detected Ralstonia solanacearum bacteria causing wilt disease with 10 times lower detection limit than ELISA in one-sixth the assay time [27]. Furthermore, multiplex microfluidic immunosensors have enabled simultaneous screening for multiple pathogens. A suspended microchannel resonator quantified mass changes due to binding between different plant virus coat proteins and their specific antibodies for multiplexed plant virus detection with high specificity. Microfluidic immunosensors thus provide faster and more sensitive detection combined with potential for highthroughput, multiplex analysis [28].

Microfluidic PCR Devices: Nucleic acid-based detection by PCR forms the gold standard for many plant diseases. However, requirements of thermal cycling instrumentation restrict widespread field adoption. Development of integrated microfluidic PCR devices has enabled simpler, portable solutions without needing complex supporting equipment. For example, an automated, battery-operated microfluidic device detected the rice blast fungus by lateral flow chromatography combined with microfluidic PCR in just 40 minutes. Using a similar strategy, citrus greening bacteria causing Huanglongbing disease was discerned in 30 minutes on a palm-sized device. These demonstrations highlight the promise of microfluidics combined with molecular testing to realize compact, easy-to-use and rapid screening devices for plant pathogens.

Table 3 highlights key examples of microfluidics/sensor integration to create practical solutions that overcome limitations of traditional diagnostic assays for detection of major plant pathogens.

Table 3. Overcoming limitations in conventional plant pathogen detection using microfluidics-integrated sensors

Pathogen	Crop	Limitation of	Microfluidic Novel
		Traditional	Approach
		Techniques	
Phytophthora	Potato	Low-throughput,	Rapid electrochemical
infestans		time-consuming	immunosensor integrated
		ELISA	with microfluidic chip
Ralstonia	Tomato	Labor-intensive	Portable microfluidic
solanacearum		culture tests	magnetoelastic
			immunosensor
Multiple	Multiple	Singular assays	Multiplex suspended
viruses		for needed each	microchannel resonators
		pathogen	
Magnaporthe	Rice	Bulky PCR machines	microfluidic Handheld
oryzae			PCR device
Candidatus	Citrus	Slow laboratory	Battery-operated
Liberibacter		analysis	microfluidic PCR detector
$B'11D$ 1	$C \times T^*$	α \cdots α	

Field Deployment of Microfluidic Sensors

A common issue plaguing current techniques is the lack of field-deployable devices which necessitates collection and transportation of samples to centralized laboratories for testing. This approach causes delays between sampling, detection and application of protective measures. Developing portable, easy-to-use, rapid detection devices is vital for timely intervention especially against highly infectious plant diseases [29]. Advances in microfluidics and lab-on-a-chip technologies coupled with smartphone connectivity have the potential to enable plant pathogen detection closer to the agriculture settings and facilitate prompt disease management decisions. For instance, a smartphone-integrated colorimetric detection system was developed to identify banana pathogens by growers themselves without requiring complex handling [30]. Such innovative solutions need to be robust, inexpensive and widely accessible at the point-of-need. Microfluidic biosensors fabricated using paper provide an attractive platform creating highly affordable diagnostic solutions requiring only a drop of sample and providing colorimetric results detectable by the naked eye. Integration of paper microfluidics with impedance sensing or molecular assays can transform field testing capacity. Despite significant promise, most microfluidic devices still remain in the development phase requiring continued research to address practical implementation challenges associated with reproducibility, standardization and scale-up which have somewhat limited technology transfer so far.

Conclusion and Future Outlook

In summary, integrating innovative sensors with well-designed microfluidic devices has generated smart platforms with powerful detection capabilities for pharmaceutical research and agricultural applications. Recent research has led to significant progress in transforming drug discovery and plant pathogen detection workflows which previously relied on legacy tools lacking in sensitivity, throughput or portability. Diverse sensing mechanisms like electrochemical, optical, masssensitive and mechanical modalities have been effectively coupled with microfluidic handling to create label-free quantified readouts of target presence by transducing biochemical events into electronic or optical signals for computational analysis. Major merits include enhanced detection limits, faster response, multiplex analysis capacity, process automation and standardization. Specifically in the drug discovery context, coupling human cell-based organs-on-chips with embedded sensors provided advanced in vitro systems that better predict clinical outcomes starting right from the initial target validation stages. Such physiologically-relevant models integrated with real-time monitoring pave the way for reliable and rapid drug screening. Adopting these microfluidic tissue models early in the pipeline also allows toxicity risk assessment improving success rates in expensive late-stage human trials. Furthermore, organs-on-chips open possibilities for developing personalized medicine by using patient-derived cells [31]. On the plant pathogen detection front, microfluidic biosensors facilitate rapid, robust and simplified molecular screening as well as immunoassays surpassing limitations of conventional PCR or ELISA which are more cumbersome needing centralized labs. These lab-onchip diagnostic tests when deployed closer to agricultural settings can enable quick intervention, containment and informed crop management strategies against infectious outbreaks saving massive losses [32].

Ongoing research continues to further enhance the capabilities and applicability of microfluidics-based sensor systems for healthcare and agriculture. Organs-on-chips are evolving with increasing complexity to better emulate human physiology by incorporating key interfaces between tissues, vascular flow, biomechanical environment and immune components. Seamless integration with various analytical modalities provides readouts to quantitatively track functional response upon exposing these mimetics to drugs, toxins or disease triggers. Expanding the variety of organ models, efforts also emphasize linking multiple organs on chip to evaluate effects of networked interactions and ADME processes upon systemic circulation models. Microfluidic organs-on-chips combined with telemedicine and electronic health record systems can possibly enable personalized medicine approaches tailored to an individual's genetic profile [33]. Additionally, coupling machine learning algorithms with drug testing on such platforms further allows intelligent,

rapid analysis to detect adverse effects early in treatment. High-throughput due to massive parallelization also facilitates precise definition of therapeutic windows for drug cocktails/combinations providing opportunities for pharmaceutical research [34].

On the plant pathogen detection side, most innovation is geared towards field deployable devices for on-site analysis eliminating delays between sampling and detection. Smartphone-based readout devices integrated with sample handling and assay steps on microfluidic chips are gaining traction to leverage mobility and connectivity. Colorimetric assays readable by naked eye minimize supporting instrumentation requirements suitable for growers [35]. Multiplexing capacity also allows simultaneous screening for multiple pathogens saving time and costs. Global spread of infectious crop diseases witnessed during recent pandemic outbreaks has further propelled the critical need for rapid, affordable and on-location testing solutions. Achieving large scale distribution requires extensive product validation through multi-site field trials and collaboration across industry partners. Efforts are also focused on further enhancing sensitivity to detect ultra-low titers at early stages, detecting resistance markers, widening target spectrum including non-culturable strains and radically simplify device operation for adoption by any end-user without extensive training. Overall integration with digital tools is poised to expand reach leveraging smartphones as readout devices. Connectivity infrastructure through wireless modules also allows remote expert consultation to strengthen preparedness and timely intervention against devastating outbreaks. Blockchain features are also being incorporated into such connected platforms to ensure data security, authentication and supply chain tracking to fight threats. Regulatory science also needs to adapt policies to cover microfluidic devices which differ vastly from traditional diagnostics improving product approval [36].

Ticrofluidics-based sensing has clearly revolutionized in vitro analysis capabilities transcending limitations of conventional methods [37]. Lab-on-chip integration continues to foster creative innovations through convergence of disciplines like microfabrication, nanotechnology, biotechnology and information technology. Transitioning more products from lab to market will widen accessibility further consolidating this technological breakthrough. Advances across both scientific and engineering domains promise continued expansion of microfluidic sensors meeting contemporary healthcare challenges and food security needs for the growing global population [38].

References

[1] F. Bouchama and M. Kamal, "Enhancing Cyber Threat Detection through Machine Learning-Based Behavioral Modeling of Network Traffic Patterns," *IJBIBDA*, vol. 4, no. 9, pp. 1–9, Sep. 2021.

- [2] R. Singh, S. Srivastava, and R. Mishra, "AI and IoT Based Monitoring System for Increasing the Yield in Crop Production," in *2020 International Conference on Electrical and Electronics Engineering (ICE3)*, 2020, pp. 301– 305.
- [3] P. Dutta, "An uncertainty measure and fusion rule for conflict evidences of big data via Dempster–Shafer theory," *International Journal of Image and Data Fusion*, vol. 9, no. 2, pp. 152–169, Apr. 2018.
- [4] N. Nguyen Thi Thanh, K. Nguyen Kim, S. Ngo Hong, and T. N. Lam, "Reply to legat, B.; Rocher, L. the limits of pairwise correlation to model the joint entropy. Comment on 'Nguyen Thi Thanh et al. Entropy correlation and its impacts on data aggregation in a wireless sensor network. Sensors 2018, 18, 3118,'" *Sensors (Basel)*, vol. 21, no. 11, p. 3729, May 2021.
- [5] B. Chen, A. Parashar, and S. Pandey, "Folded floating-gate CMOS biosensor for the detection of charged biochemical molecules," *IEEE Sensors Journal*, vol. 11, no. 11, pp. 2906–2910, 2011.
- [6] I. Kiselev *et al.*, "Erratum: Kiselev, I., et al. On the temporal stability of analyte recognition with an E-nose based on a metal oxide sensor array in practical applications. Sensors 2018, 18, 550," *Sensors (Basel)*, vol. 19, no. 16, p. 3525, Aug. 2019.
- [7] A. Nassar and M. Kamal, "Machine Learning and Big Data Analytics for Cybersecurity Threat Detection: A Holistic Review of Techniques and Case Studies," *Intelligence and Machine Learning …*, 2021.
- [8] C. M. Legner, G. L. Tylka, and S. Pandey, "Robotic agricultural instrument for automated extraction of nematode cysts and eggs from soil to improve integrated pest management," *Scientific Reports*, vol. 11, no. 1, p. 3212, 2021.
- [9] T. Ye, B. Wang, P. Song, and J. Li, "Correction: Ye, T.; Et al. Automatic railway traffic object detection system using feature fusion refine neural network under shunting mode. Sensors 2018, 18, 1916," *Sensors (Basel)*, vol. 19, no. 14, p. 3044, Jul. 2019.
- [10] Sensors Editorial Office, "Acknowledgement to reviewers of sensors in 2016," *Sensors (Basel)*, vol. 17, no. 12, p. 128, Jan. 2017.
- [11] A. Nassar and M. Kamal, "Ethical Dilemmas in AI-Powered Decision-Making: A Deep Dive into Big Data-Driven Ethical Considerations," *IJRAI*, vol. 11, no. 8, pp. 1–11, 2021.
- [12] J. N. Saldanha, A. Parashar, S. Pandey, and J. A. Powell-Coffman, "Multiparameter behavioral analyses provide insights to mechanisms of cyanide resistance in Caenorhabditis elegans," *toxicological sciences*, vol. 135, no. 1, pp. 156–168, 2013.
- [13] T. Kong, N. Backes, U. Kalwa, C. Legner, G. J. Phillips, and S. Pandey, "Adhesive tape microfluidics with an autofocusing module that incorporates CRISPR interference: applications to long-term bacterial antibiotic studies," *ACS sensors*, vol. 4, no. 10, pp. 2638–2645, 2019.
- [14] M. Muniswamaiah, T. Agerwala, and C. C. Tappert, "Context-aware query performance optimization for big data analytics in healthcare," in *2019 IEEE High Performance Extreme Computing Conference (HPEC-2019)*, 2019, pp. $1 - 7$.
- [15] E. Kutafina, D. Laukamp, R. Bettermann, U. Schroeder, and S. M. Jonas, "Correction: Kutafina, E.; Laukamp, D.; Bettermann, R.; Schroeder, U.; Jonas, S.m. wearable sensors for eLearning of manual tasks: Using forearm EMG in hand hygiene training. Sensors 2016, 16, 1221," *Sensors (Basel)*, vol. 19, no. 21, p. 4792, Nov. 2019.
- [16] M. Muniswamaiah, T. Agerwala, and C. C. Tappert, "Federated query processing for big data in data science," in *2019 IEEE International Conference on Big Data (Big Data)*, 2019, pp. 6145–6147.
- [17] U. Kalwa, C. Legner, E. Wlezien, G. Tylka, and S. Pandey, "New methods of removing debris and high-throughput counting of cyst nematode eggs extracted from field soil," *PLoS One*, vol. 14, no. 10, p. e0223386, 2019.
- [18] Y. Li, H. Wang, W. Zhu, S. Li, and J. Liu, "Structural stability monitoring of a physical model test on an underground cavern group during deep excavations using FBG sensors," *Sensors (Basel)*, vol. 15, no. 9, pp. 21696– 21709, Aug. 2015.
- [19] M. Kamal and T. A. Bablu, "Machine Learning Models for Predicting Clickthrough Rates on social media: Factors and Performance Analysis," *IJAMCA*, vol. 12, no. 4, pp. 1–14, Apr. 2022.
- [20] A. Tripathy, S. Pramanik, J. Cho, J. Santhosh, and N. A. A. Osman, "Role of morphological structure, doping, and coating of different materials in the sensing characteristics of humidity sensors," *Sensors (Basel)*, vol. 14, no. 9, pp. 16343–16422, Sep. 2014.
- [21] Z. Njus *et al.*, "Flexible and disposable paper-and plastic-based gel micropads for nematode handling, imaging, and chemical testing," *APL bioengineering*, vol. 1, no. 1, 2017.
- [22] S.-M. Yu, U.-S. Jeong, H. K. Lee, S. H. Baek, S. J. Kwon, and Y. H. Lee, "Disease occurrence in transgenic rice plant transformed with silbene synthase gene and evaluation of possible horizontal gene transfer to plant pathogens," *Sigmulbyeong Yeongu*, vol. 20, no. 3, pp. 189–195, Sep. 2014.
- [23] I. Stergiopoulos and T. R. Gordon, "Cryptic fungal infections: the hidden agenda of plant pathogens," *Front. Plant Sci.*, vol. 5, p. 506, Sep. 2014.
- [24] Sensors Editorial Office, "Acknowledgment to reviewers of sensors in 2021," *Sensors (Basel)*, vol. 22, no. 3, p. 1052, Jan. 2022.
- [25] A. Q. Beeman, Z. L. Njus, S. Pandey, and G. L. Tylka, "Chip technologies for screening chemical and biological agents against plant-parasitic nematodes," *Phytopathology*, vol. 106, no. 12, pp. 1563–1571, 2016.
- [26] X. Li, H. Zhao, and X. Chen, "Screening of marine bioactive antimicrobial compounds for plant pathogens," *Mar. Drugs*, vol. 19, no. 2, p. 69, Jan. 2021.
- [27] N. Yahata *et al.*, "Anti-Aβ drug screening platform using human iPS cellderived neurons for the treatment of Alzheimer's disease," *PLoS One*, vol. 6, no. 9, p. e25788, Sep. 2011.
- [28] J. Song and A. F. Bent, "Microbial pathogens trigger host DNA double-strand breaks whose abundance is reduced by plant defense responses," *PLoS Pathog.*, vol. 10, no. 4, p. e1004030, Apr. 2014.
- [29] X. Ding, Z. Njus, T. Kong, W. Su, C.-M. Ho, and S. Pandey, "Effective drug combination for Caenorhabditis elegans nematodes discovered by outputdriven feedback system control technique," *Science advances*, vol. 3, no. 10, p. eaao1254, 2017.
- [30] A. M. Lerario and G. D. Hammer, "Drug repurposing using high-throughput screening identifies a promising drug combination to treat adrenocortical carcinoma," *Oncotarget*, vol. 9, no. 70, pp. 33245–33246, Sep. 2018.
- [31] R. Finger, S. M. Swinton, and N. El Benni, "Precision farming at the nexus of agricultural production and the environment," *Annual Review of*, 2019.
- [32] W. J. Vlietstra, R. Vos, A. M. Sijbers, E. M. van Mulligen, and J. A. Kors, "Using predicate and provenance information from a knowledge graph for drug efficacy screening," *J. Biomed. Semantics*, vol. 9, no. 1, p. 23, Sep. 2018.
- [33] T. Kong, R. Brien, Z. Njus, U. Kalwa, and S. Pandey, "Motorized actuation system to perform droplet operations on printed plastic sheets," *Lab on a Chip*, vol. 16, no. 10, pp. 1861–1872, 2016.
- [34] S. Sari *et al.*, "Antifungal screening and in silico mechanistic studies of an inhouse azole library," *Chem. Biol. Drug Des.*, vol. 94, no. 5, pp. 1944–1955, Sep. 2019.
- [35] The PLOS Pathogens Staff, "Correction: Microbial pathogens trigger host DNA double-strand breaks whose abundance is reduced by plant defense responses," *PLoS Pathog.*, vol. 10, no. 6, p. e1004226, Jun. 2014.
- [36] C. K. Choi and H. H. Yoo, "Precision fault diagnosis procedure for a structural system having a defect employing Hidden Markov Models," *Int. J. Precis. Eng. Manuf.*, vol. 15, no. 8, pp. 1667–1673, Aug. 2014.
- [37] J. A. Carr, R. Lycke, A. Parashar, and S. Pandey, "Unidirectional, electrotactic-response valve for Caenorhabditis elegans in microfluidic devices," *Applied Physics Letters*, vol. 98, no. 14, 2011.
- [38] T. Wollenberg and J. Schirawski, "Comparative genomics of plant fungal pathogens: the Ustilago-Sporisorium paradigm," *PLoS Pathog.*, vol. 10, no. 7, p. e1004218, Jul. 2014.